

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/234161503>

The vascular biology of macrophage migration inhibitory factor (MIF). Expression and effects in inflammation...

Article in *Thrombosis and Haemostasis* · January 2013

DOI: 10.1160/TH12-11-0831 · Source: PubMed

CITATIONS

43

READS

87

3 authors:



[Yaw Asare](#)

Ludwig-Maximilians-University of Munich

16 PUBLICATIONS 128 CITATIONS

[SEE PROFILE](#)



[Martin M N Schmitt](#)

Ludwig-Maximilians-University of Munich

14 PUBLICATIONS 209 CITATIONS

[SEE PROFILE](#)



[Jürgen Bernhagen](#)

Ludwig-Maximilians-University of Munich, Kli...

234 PUBLICATIONS 12,438 CITATIONS

[SEE PROFILE](#)

The vascular biology of macrophage migration inhibitory factor (MIF)

Expression and effects in inflammation, atherogenesis and angiogenesis

Yaw Asare¹; Martin Schmitt^{2,3}; Jürgen Bernhagen¹

¹Institute of Biochemistry and Molecular Cell Biology, RWTH Aachen University, Aachen, Germany; ²Institute of Molecular Cardiovascular Research (IMCAR), RWTH Aachen University, Aachen, Germany; ³Cardiovascular Research Institute Maastricht (CARIM), Maastricht University, The Netherlands

Summary

Macrophage migration inhibitory factor (MIF) is a pleiotropic cytokine with chemokine-like functions. MIF is a critical mediator of the host immune and inflammatory response. Dysregulated MIF expression has been demonstrated to contribute to various acute and chronic inflammatory conditions as well as cancer development. More recently, MIF has been identified as an important pro-atherogenic factor. Its blockade could even aid plaque regression in advanced atherosclerosis. Promotion of atherogenic leukocyte recruitment processes has been recognised as a major underlying mechanism of MIF in vascular pa-

thology. However, MIF's role in vascular biology is not limited to immune cell recruitment as recent evidence also points to a role for this mediator in neo-angiogenesis / vasculogenesis by endothelial cell activation and endothelial progenitor cell recruitment. On the basis of introducing MIF's chemokine-like functions, the current article focusses on MIF's role in vascular biology and pathology.

Keywords

Chemokine, monocyte/macrophage, cardiovascular disease, atherosclerosis, (neo-)angiogenesis/vasculogenesis

Correspondence to:

Univ.-Prof. Dr. rer. nat. Jürgen Bernhagen
Institute of Biochemistry and Molecular Cell Biology
RWTH Aachen University, Pauwelsstrasse 30, D-52074 Aachen, Germany
Tel.: +49 241 80 88 840/31/41, Fax: +49 241 80 82 427
E-mail: jbernhagen@ukaachen.de

Received: November 18, 2012

Accepted after minor revision: December 3, 2012

Prepublished online: January 17, 2013

doi:10.1160/TH12-11-0831

Thromb Haemost 2013; 109: ■■■

Introduction: chemokines and vascular biology

The processes governing vascular homeostasis, vascular repair after acute injury and vascular remodelling during chronic disease are controlled and driven by a plethora of factors. Among them chemokines play a pivotal role at all levels of regulation.

Chemokines are small chemoattractant cytokines which have a molecular weight of 8-12 kDa. Chemokines exert a broad variety of functions in physiology and pathophysiology. In the context of the current review article, we will focus on their role in leukocyte chemotaxis, extravasation, as well as augmentation and/or attenuation of angiogenesis (1-3). Based on the arrangement of four conserved cysteine residues, chemokines are divided into four families (CC, CXC, CX3C, and XC) (4). Numerous examples of key roles of chemokines in vascular function, atherogenesis and vascular repair exist. A joint role for several of these molecular players can easily be rationalised, if one considers that the different steps of leukocyte recruitment process, i.e. rolling, firm adhesion, and transmigration are controlled by functionally specialised chemokines, which act in a sequential and cooperative manner. However, in the context of this review, we can only refer to some excellent previous review articles covering the aspects of a balance between chemokine redundancy and cooperativity in both the maintenance of vascular homeostasis and vascular pathology (3, 5-8).

Chemokine structure and function may be modulated by di-, tetrameric or perhaps even higher-order interactions. These inter-

actions are usually homomeric (for a comprehensive review please see [9]), but chemokine function can also be modulated by heteromerisation, e.g. between CC and CXC chemokines. This may serve to promote chemokine/receptor interactions. An example for the latter is the heterodimerisation between CXCL4/PF4 and CCL5/RANTES, which takes place in α -granules of human platelets deposited at the glycosaminoglycan surface of endothelial cells *in vivo*. *In vitro* and *in vivo* studies revealed a pathophysiological function of such chemokine heterodimers (10, 11). In fact, von Hundelshausen et al. demonstrated that heterodimerisation of CCL5 and CXCL4 enhances CCL5-mediated monocyte recruitment, while Koenen *et al.* additionally identified peptidic inhibitors specifically interrupting the CCL5/CXCL4 interface and inhibiting vascular lesion formation in atherosclerotic *ApoE*^{-/-} mice (10, 11).

A subfamily of the CXC chemokine family is known as the ELR+ chemokines and is defined by a glutamic acid-leucine-arginine (ELR)-motif near the CXC sequence. In contrast to non-ELR chemokines (ELR- CXC chemokines), chemokines from the ELR+ subfamily are potent inducers of physiological and pathological angiogenesis. They play important roles in diseases like cancer, fibroproliferative disorders and chronic inflammation like atherosclerosis (12-14). A representative example for an ELR+ chemokine is CXCL8/interleukin (IL-8). Thus, besides being a potent neutrophil chemoattractant, CXCL8 also plays an important role in neovascularisation in human non-small cell lung cancer (15), human gastrointestinal cancers (16) and human ovarian and pros-

tate cancer (17), where the CXCL8-CXCR2 axis regulates tumour angiogenesis accompanied by correlative reduction or enhancement of tumour growth. Further evidence for CXCL8-mediated angiogenesis in diseased tissue was found in human coronary artery plaques, where CXCL8 has been shown to be overexpressed (18). This might point to CXCL8-mediated growth of intra-plaque vessels and plaque destabilisation (19).

As an example for a prominent CC chemokine, the CCL2/CCR2 ligand receptor axis is important in monocyte chemoattraction and transendothelial migration into areas of vascular inflammation. CCL2 expression has been shown in atherosclerotic lesions, likely exacerbating lesion progression through extensive monocyte attraction and also through triggering firm monocyte adhesion to the inflamed endothelium (20, 21). A deletion or attenuation of CCL2 expression in atherosclerotic mouse models resulted in decreased lesion formation (22, 23).

In addition to the four canonical chemokine classes, a group of molecules sharing functional similarities with chemokines has emerged as a fifth subclass. This class has been referred to as 'chemokine-like function' (CLF) chemokines, non-canonical chemokines, or micro-chemokines. By definition, the CLF family of chemokines encompasses certain inflammatory and immune proteins that exhibit chemokine-like functions such as chemotactic properties or leukocyte arrest-promoting effects but which neither formally carry the typical N-terminal cysteine motif of the classical chemokines nor the chemokine fold. Most members of this functional family have been found to act as non-canonical ligands for classical chemokine receptors. CLF chemokines thus further expand the degree of redundancy and promiscuity in chemokine/chemokine receptor interactions, with consequences for angiogenesis regulation, as discussed below.

Examples are a cleavage fragment of tyrosyl tRNA synthetase (TyrRS) which has been shown to act as a non-canonical CXCR1 ligand via the presence of an ELR-like motif (24). The N-terminal 'mini-TyrRS' domain has proangiogenic properties and induces neutrophil chemotaxis through interaction with CXCR1 but not CXCR2 (24, 25). Interestingly, the ELR motif is only exposed and available for interaction with CXCR1 in the cleaved fragment (26). Similarly, autoantigenic aminoacyl-RS, released under apoptotic conditions, have leukocyte recruitment properties by triggering CC receptors. Both histidyl-RS and its N-terminal fragment are chemoattractants for several CCR5-expressing immune cells. Asparaginyl-RS interacts with CCR3 (27). For these aminoacyl-RS, the presence of specific surface charge distributions has been suggested to mediate chemokine receptor usage. Furthermore, the human antimicrobial peptides β -defensin-1 and -2 were identified as non-cognate ligands for CCR6, mediating chemotaxis (28). Again, although the sequence similarity between the β -defensins and CCL20 is limited, it appears that a cluster of cationic amino acids and shared electrostatic charge patterns account for the overlap in chemotactic activities. Promiscuity and mimicry mechanisms not only can be found endogenously in the host, but there are several examples of parasite or viral chemokine mimicry, encompassing both mimicry of classical host chemokine structures and CLF-type mimicry mechanisms (29, 30). Most prominently,

HIV-1 capsid protein gp120 interacts with host CXCR4 (and CCR5) to direct leukocyte infection (29, 31). The nuclear protein high-mobility-group binding protein 1 (HMGB1) has been shown to exert numerous extracellular inflammatory functions. HMGB1 signals through several receptors (32, 33) and most recently, heteromeric complex formation between HMGB1 and CXCL12 was identified to mediate HMGB1 chemokine activities through CXCR4 (34).

In this review, we will focus on macrophage migration inhibitory factor (MIF), one of the first cytokines to be discovered (35) and another recent addition to the CLF family of chemokines (30, 36). In the next chapters, we will briefly outline both the physiology and the pathophysiologic roles of MIF in inflammatory and vascular disease, including underlying mechanisms such as its non-cognate interactions with the chemokine receptors CXCR2 and CXCR4. With regard to MIF's mechanisms of action, we will solely focus on aspects relating to vascular biology and pathophysiology.

MIF: structure, mechanism of action, and role in inflammatory disease

MIF is an evolutionarily-conserved protein that is abundantly expressed in humans and non-primate mammals. In addition to its functions as cytokine/chemokine and angiogenic factor (see below), it has been suggested that MIF also has anti-oxidative intracellular effects. The MIF structure is unique among cytokines. MIF consists of 114 amino acids and has a molecular weight of 12.5 kDa. The three-dimensional structure of human MIF shows that MIF crystallises as a trimer of three identical subunits, but studies at more physiological concentrations imply that the monomer may have crucial functions *in vivo* as well. Despite its wide tissue distribution, the secretion of MIF is tightly regulated, with relevant triggers such as hypoxia/ischaemia or oxidised low-density lipoprotein (oxLDL) of particular importance for this review article. Moreover, MIF expression is strongly up-regulated in several disease conditions most importantly in vascular pathology and tumourigenesis (37-39).

CD74, the membrane-expressed form of invariant chain (Ii) and an MHC class II chaperone, was identified as the first MIF plasma membrane receptor (40). CD74 expression is typically restricted to class II-positive cells, but under inflammatory conditions as well as in several tumour cell types, CD74 can be up-regulated, even in the absence of class II expression. MIF binds to CD74 by high affinity interaction in the nanomolar range, but signalling additionally requires the recruitment of signalling-competent co-receptors such as CD44 or CXCR2 and CXCR4 (see below). CD74 alone mediates MIF binding, but MIF-induced MAPK signalling requires the coexpression of CD44. MIF signalling through CD74/CD44 also involves the serine phosphorylation of the cytoplasmic tails of CD74 and CD44 and the recruitment of a Src-type tyrosine kinase (41). An architectural similarity between the MIF monomer and the CXCL8 dimer instigated biochemical investigations to probe potential interactions between MIF and CXCR2.

Receptor binding studies then revealed that MIF engages in a non-cognate, high affinity interaction with CXCR2 (42). CXCR2 had been known to be promiscuous previously, as it also is the receptor for six other ELR+ CXC chemokines including CXCL8. This as well as numerous functional data then suggested that MIF belongs to the family of CLF chemokines. The signal transduction pathways triggered by MIF/CXCR2 have not been studied systematically, but initial evidence shows that MIF binding to CXCR2 leads to G_i coupling and can elicit calcium transients. A limited screen of other chemokine receptors as well as an observed chemotactic effects of MIF on T cells, which do not express CXCR2 and only low levels of CD74, unraveled yet another MIF/chemokine receptor interaction, i.e. that with CXCR4. MIF/CXCR4 interaction is less affine than that between MIF and CXCR2 but still in the nanomolar range. Further research into the receptors of MIF revealed that CD74 forms heteromeric complexes with either CXCR2 or CXCR4 (42, 43).

During inflammation (44), endothelial cells do not only get activated but also adjust their phenotypes to the inflammatory response (45). Activation of endothelial cells (ECs), however, contributes to both acute and chronic inflammatory diseases such as sepsis, inflammatory bowel disease, rheumatoid arthritis, inflammatory lung disease, and atherosclerosis (46). There is ample evidence now that MIF is a key mediator of all of these disease conditions (37, 38, 47-49). Thus, MIF has been proven to play a pivotal role in the pathogenesis of both acute and chronic inflammatory diseases. Two prominent examples are sepsis and rheumatoid arthritis.

First evidence implicating MIF in systemic infection and sepsis dates back two decades ago when pituitary-derived MIF was found to contribute to serum levels of MIF in the post-acute phase of endotoxaemia. Employing a mouse model of endotoxic shock, MIF was found to promote lethal endotoxaemia in mice (50). Indeed, several and diverse models of septic shock have demonstrated the crucial role of MIF in mediating sepsis (51-53).

Rheumatoid arthritis, a systemic chronic inflammatory disorder of the joints, is characterised by key pathological events including diapedesis of leukocytes and the active participation of cytokines such as tumour necrosis factor (TNF). MIF has been extensively described to play a role in rheumatoid arthritis by e.g. inducing the secretion of CCL2 and to promote TNF production to amplify leukocyte recruitment at yet another level (38, 54). Of note, polymorphisms in the *MIF* gene functionally enhancing the transcriptional activity of MIF have been linked to increased disease severity of rheumatoid arthritis and other inflammatory conditions (38, 55).

MIF in the vasculature

Vascular endothelial cells exhibit a profound heterogeneity and organ specificity in terms of their phenotype and protein expression patterns. Depending on the vessel or tissue they inhabit, ECs are either strictly continuous with tight junctions, e.g. to maintain the blood-brain barrier or discontinuous in the case of the liver to allow for maximal fluid exchange. On the other hand,

ECs lining the glomerulus are strongly fenestrated to allow for optimal filtration results. In large arteries, an additional requirement to resist high pressure and shear flow is found. ECs express various molecules that have been described to be pivotal in the pathogenesis of numerous vascular diseases such as atherosclerosis and angiogenesis. Among these molecules are various adhesion molecules, such as vascular cell adhesion molecules (VCAMs), intercellular adhesion molecules (ICAMs), selectins, and junctional adhesion molecules (JAMs).

The expression levels of MIF in human ECs of both microvascular and large artery origin have been shown to be upregulated upon treatment with oxLDL (56) or thrombin (57) in a time- and dose-dependent manner, suggesting a role for MIF in the vasculature. MIF released upon oxLDL stimulation contributes to atherogenic leukocyte recruitment (56, 58) (► Figure 1). Moreover, exposing human ECs to hypoxia led to a release of substantial amounts of MIF that was found to participate in the recruitment and migration of endothelial progenitor cells (59). The expression of MIF in the vasculature extends beyond the endothelial layer as vascular smooth muscle cells (VSMCs) have also been shown to express low levels of MIF (56); moreover, MIF expression was also observed to be upregulated by oxLDL in VSMCs (60). Importantly, VSMCs do not only express MIF but also migrate towards exogenous MIF after 6 hours (h) of incubation (61) (► Figure 1).

MIF signalling in the vasculature has also been pursued. Subsequent to showing MIF-induced expression of ICAM-1 in ECs (62), Cheng et al. reported the expression of VCAM-1, E-selectin, ICAM-1 and CCL2 to be MIF-dependent. Indeed, TNF-induced leukocyte rolling and adhesion to MIF-depleted human umbilical vein endothelial cells (HUVECs) were impaired, an observation that was attributed to a reduction of p38 MAPK activation, resulting in reduced expression of chemokines and adhesion molecules (63). In addition, MIF was found to upregulate mononuclear cell adhesion molecules such as VCAM-1 and ICAM-1 in a Src-, nuclear factor- κ B (NF- κ B), and phosphatidylinositol-3-kinase (PI3K)-dependent manner (64). The above findings are in line with a previous report where oxLDL- and MIF-induced monocyte arrest on HAoEC was abrogated by anti-MIF neutralising antibody (58). Moreover, MIF blockade leads to reduced VSMC proliferation and neointimal thickening (65), while at later stages during the atherogenic process MIF contributes to plaque instability (58). Thus, it is not surprising that there is extensive literature on the functional role of vascular MIF in the pathogenesis of vascular disease like atherosclerosis or neointimal growth after vascular injury.

MIF in atherosclerosis

Inflammatory processes are key contributors to the pathogenesis of atherosclerosis (66). The proinflammatory chemokine-like cytokine MIF has been broadly implicated in atherogenesis (39) (► Figure 1). Expression of MIF in developing atherosclerotic plaques in humans was minimal in ECs and smooth muscle cells (SMCs) in non-lesion-associated areas but was upregulated in ECs, SMCs, macrophages and T cells upon atheroprogession suggesting a role for MIF in plaque instability (56, 67). Interestingly,

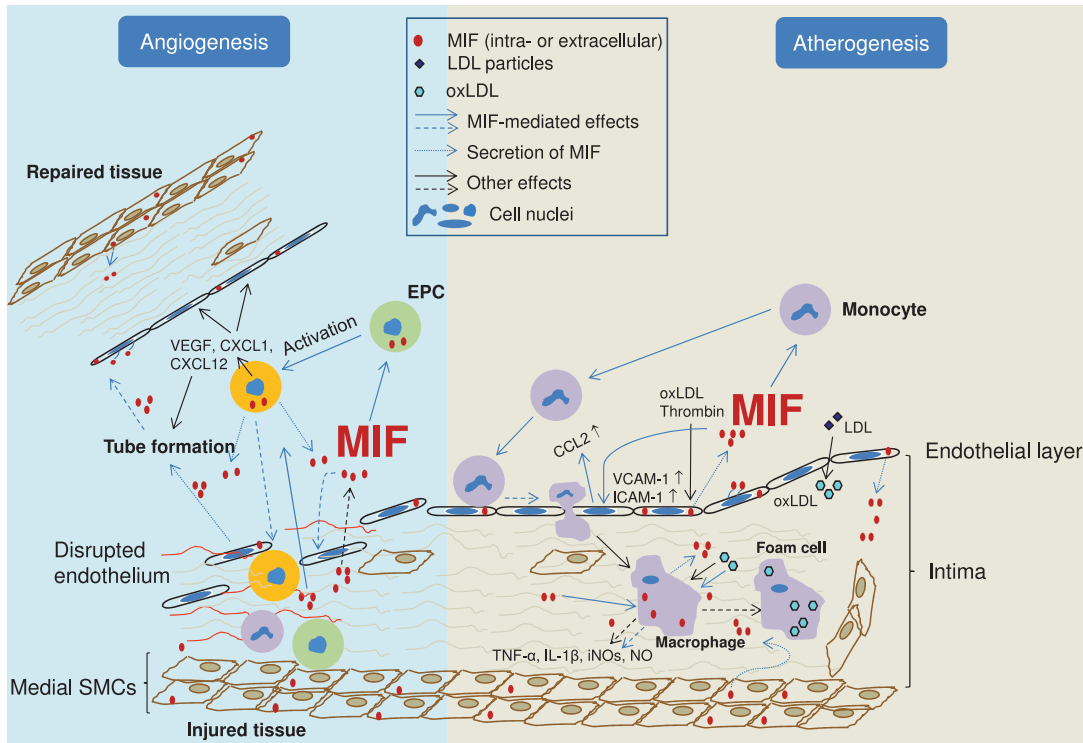


Figure 1: Expression and functional role of vascular MIF in angiogenesis and atherosclerosis. Left (angiogenesis): After tissue injury, MIF along with other chemokines such as CXCL12, CXCL1, and VEGF is released to activate and recruit EPCs and also monocytes to the site of injury where they are embedded into tube structures. Importantly, MIF drives tube formation and the formation of new vessels that follows. EPCs carry angiogenic factors ('cargo') themselves. Right (atherosclerosis): MIF expressed by ECs and macrophages in the atherosclerotic plaque is upregulated upon stimulation with inflammatory and atherogenic mediators such as oxLDL and thrombin. MIF induces the expression of chemokines (CCL2) and adhesion molecules (VCAM-1, ICAM-1) which regulates the recruitment and adhesion of monocytes to the surface of the endothelium. Alternatively, MIF may employ its

chemokine receptors CXCR2 and CXCR4 (expressed on recruited monocytes and T cells, respectively; receptors not shown) in exerting its chemokine-like functions. After transmigration into the subendothelial space, monocytes subsequently differentiate into macrophages. MIF and other pro-atherogenic factors (latter not shown) stimulate these macrophages to secrete TNF- α , IL-1 β , iNOS and NO, enhancing the inflammatory milieu in the lesion. Additionally, MIF potentiates foam cell formation by accelerating macrophage oxLDL uptake, thereby enhancing lesion formation. VSMCs express and also migrate towards MIF and this may contribute to plaque stability, although long term exposure to MIF has also been shown to inhibit PDGF-BB-induced VSMCs migration and MMP upregulation which points in the direction of plaque destabilisation by MIF (for references see text).

an interaction between MIF and CSN5/JAB1, a negative regulator of NF- κ B which is a key transcription factor involved in the inflammatory and immune processes associated with atherosclerosis (68, 69), was revealed in human atherosclerotic lesions (56). As a negative regulator of CSN5, MIF might synergise with JAB1 in regulating NF- κ B-driven inflammatory and immune signalling in atherosclerosis (70).

Atherosclerosis may be preceded by vascular injury and MIF has been shown to regulate the biological response to the injured tissue. In a carotid artery injury model of atherosclerosis-susceptible mice, MIF was shown to potentiate neointimal thickening by promoting the accumulation of inflammatory cells in the neointima and the proliferation of medial and intimal cells (65). Indeed, neutralising MIF resulted in a reduction of neointimal macrophage content and an increase in SMCs and collagen type I content in the neointimal lesions in a model of vascular injury-induced accelerated lesion formation. This reduction in neointimal macrophage content was attributed to impaired monocyte recruitment as exogenous MIF increased the number of monocytes adhering to

HAoECs, an effect that was dampened by neutralising MIF mAbs (58). The notion that MIF is critical in the development of atherosclerosis was corroborated by Mif gene-inactivation in *Ldlr*^{-/-} mice. These mice showed impaired atherogenic diet-induced lesion initiation and progression when compared with corresponding wild-type mice. The ability of MIF to induce proliferation of SMCs which has previously been observed mainly by using mAb was confirmed by this genetic study (71). In a spontaneous atherogenic *Apoe*^{-/-} mouse model, MIF was found to be elevated at the aortic wall and blocking aortic MIF with anti-MIF neutralising antibody led to a reduction in intimal macrophage content and inflammatory mediators (72). Notably, treatment with a blocking antibody that targets MIF even resulted in atherosclerotic plaque regression and a more stable plaque phenotype in diet-induced atherosclerotic *Apoe*^{-/-} mice. This observation was reasserted by the finding that MIF mediates atherogenic monocyte and T cell recruitment *in vivo* by engaging its receptors CXCR2 and CXCR4, respectively (► Figure 1). Dual action of MIF through CXCR2 and CXCR4 also may explain why anti-MIF

blockade was more efficacious in the clinically highly relevant regression model than treatment with anti-CXCL1 or anti-CXCL12 (42). Indeed, a MIF N-loop-derived peptide that disrupts the interaction between MIF and CXCR2 blocked MIF-induced leukocyte adhesion in carotid arteries *in vivo* (73).

Taken together, data from several groups using either neutralising antibody or genetic deletion underpins the pathogenic role of MIF in promoting atherogenic changes in the arterial vessel wall. Importantly, the proatherogenic role of MIF was confirmed in epidemiologic studies where single nucleotide polymorphisms (SNPs) in the human *MIF* gene were identified as a risk factor for coronary heart disease, showing an association of a haplotype containing the rs755622C allele, which has been reported before to increase the susceptibility for various other proinflammatory conditions (74), and showing that the GG genotype of the MIF SNP rs1007888 was associated with myocardial infarction (MI) in Czech female patients (75). Moreover, Makino et al. showed that high plasma levels of MIF are associated with adverse long-term outcome in patients with stable coronary artery disease and impaired glucose tolerance or type 2 diabetes mellitus (76). Similarly, Müller et al. found that MIF expression is enhanced in acute coronary syndromes (ACS), that it is associated with various markers of the inflammatory response, that it correlates with the extent of cardiac necrosis marker release after percutaneous intervention and that it is increased in ACS patients with respective lesions (77).

Role of MIF in (neo-)angiogenesis / vasculogenesis

MIF's role in atherogenesis has been extensively studied (see above); however, its roles in the processes controlling physiologic and pathophysiologic angiogenesis are less well understood.

Angiogenesis is the growth of blood vessels from a pre-existing vasculature. It occurs throughout life and is an important component of different physiological and pathophysiological conditions such as wound healing and pregnancy (78). Regulation of angiogenesis is achieved by balancing angiogenic and angiostatic triggers. If not properly controlled, angiogenesis can promote to tumour growth, rheumatic arthritis, and retinopathies (78). The therapeutic value of angiogenesis has become of great interest. Inhibiting or decreasing angiogenesis possesses therapeutic potential in treating cancer and rheumatic arthritis, while stimulation of angiogenesis can be helpful in ischaemic heart disease, peripheral arterial disease, and wound healing responses by increasing reperfusion of the tissue. Moreover, vasculogenesis, a process formerly considered to be restricted to the *de novo* formation of vascular structures from mesenchymal angioblasts in early embryonic vascular development (79, 80), has now also been discussed to occur postnatally, where it is triggered by endothelial progenitor cells (EPCs¹) (81–84), opening-up promising novel therapeutic avenues as well (85, 86).

Oxygen plays a crucial role in controlling angiogenesis. Hypoxic conditions stimulate vessel growth by activation of ECs (i.e. proliferation and migration). The cellular response to hypoxia is

mediated via up-regulation of hypoxia-inducible transcription factors (HIFs), most prominently HIF-1 α . HIFs upregulate the transcription of numerous genes, thereby affecting endothelial cell growth, SMC recruitment, and leukocyte attraction. HIF-1 α is the best characterised inducer of the expression of vascular endothelial growth factor (VEGF), the major endothelial growth factor in angiogenesis and a key trigger of angiogenesis. Hypoxia-triggered VEGF production and the subsequent increase in oxygen supply following newly formed vessel growth reciprocally regulate each other to restore homeostasis following limited blood supply (86–88).

MIF was first implicated in angiogenesis some 14 years ago, when Chesney et al. found that MIF blockade by a neutralising antibody reduces tumour vascularisation and tumour growth in a murine model of B-cell lymphoma (89). Although several distinct mechanisms have been suggested to underlie MIF's potent pro-tumorigenic capacity, a role for MIF in tumour angiogenesis was confirmed in a model of colon adenocarcinoma formation, in which blockade of MIF led to reduced microvessel formation (90). Moreover, MIF expression is seen in non-small-cell lung cancer, a tumour entity in which MIF now has been firmly established as a crucial player (91–95). MIF expression occurs in association with angiogenic CXC chemokines and increased vessel density (70). As discussed above, MIF is a non-cognate ligand for CXCR2, the cognate receptor for angiogenic CXCL8, and MIF also promiscuously engages CXCR4, the cognate receptor for CXCL12. Although CXCL12 is an ELR- CXC chemokine, both CXCR2 and CXCR4 have been found involved in numerous pro-angiogenic effects in various models of postnatal angiogenesis, including post-ischaemic adaption (71–73), underscoring the notion that the chemokine receptors of MIF could be critical in mediating MIF-driven pro-angiogenic responses, although direct evidence from knockout mouse models is yet missing for this assumption (► Figure 1).

As for angiogenic factors such as VEGF, MIF expression was also identified to be regulated by HIF-1 α activity, as demonstrated in lung tissue, ECs, hepatic stellate cells, and VSMCs (59, 96–98). A number of studies have since confirmed MIF's pro-angiogenic properties and explored the underlying molecular and cellular mechanisms. MIF mediates EC migration and tube formation in matrigel assays and induces angiogenesis in matrigel plugs and the cornea angiogenesis assay (► Figure 1). These effects rely on mitogen-activated protein kinase (MAPK) and PI3K signalling, activities known to be triggered by MIF (42, 99–101). Accordingly,

1 Endothelial progenitor cells (EPCs) were initially considered to represent a single entity of progenitor cells capable of supporting post-natal *de novo* blood vessel formation and have been assigned a crucial role in neo-angiogenesis. However, since their discovery in 1997 their phenotype has been refined and become more complex. In fact, EPCs exhibit different characteristics: (i) the so-called early outgrowth EPCs (EOCs) are derived from circulating CD34-positive mononuclear cells and additionally express CD45 and CD14; they exert enhanced adhesion properties but fail to proliferate *in vitro*; (ii) the so-called late outgrowth EPCs (LOCs), lack hematopoietic markers but have the ability to proliferate. Both subtypes respond to angiogenic stimuli, express CD31 and secrete angiogenic factors such as VEGF and angiogenic cytokines/chemokines by themselves (79–82).

MIF could be detected in the tumour-associated neovasculature and neointima following vascular injury in pro-atherogenic mouse models (58, 89, 99).

Tissue repair after MI heavily relies on neoangiogenesis of the infarcted area. Chemokines and their receptors play important roles during these 'repair' processes. Both exacerbating and pro-healing responses occur. For example, the genetic absence of CC chemokine receptor Ccr1 in a corresponding mouse model reduces functional impairment and structural remodelling after MI due to an abrogated early inflammatory recruitment of neutrophils and improved tissue healing including vessel regeneration (102). Interestingly, the CXCL12/MIF receptor CXCR4 was recently found to have a dual role in neo-angiogenesis after MI. CXCR4 was found to play a crucial role in endogenous remodelling processes after MI, contributing to inflammatory/progenitor cell recruitment and neovascularisation, whereas its deficiency limits infarct size and causes adaptation to hypoxic stress (103).

MIF has now been amply implicated as a protective factor in MI-ischaemia/reperfusion (I/R) injury. Although the protective mechanisms involved have been suggested to span from AMP kinase activation to promotion of anti-oxidative pathways (104-107), it is likely that MIF-driven angiogenic/vasculogenic processes are equally important. Indeed, MIF secreted from ECs by hypoxic stimulation has been identified to promote EPC chemotaxis in a CXCR4- (59) and CXCR2- (D. Simons and J. Bernhagen, unpublished observations) dependent manner (► Figure 1). This finding was intriguing, because previously CXCL12 was considered the main protagonist in driving EPC recruitment following hypoxic/ischaemic triggers (108, 109). However, these seemingly divergent findings may be readily reconciled if one considers the kinetics of chemokine production. Hypoxia-stimulated HUVECs and human aortic endothelial cells (HAoECs) did not secrete detectable CXCL12 levels within an early time window of 2 h, when MIF was predominantly secreted, peaking at 60 minutes (59). This nicely coincides with the findings by Ceradini et al. reporting that CXCL12 production from hypoxically challenged HUVECs and ischaemic tissue *in vivo* occurred in a time interval of 6-24 h and correlated subsequent EPC trafficking *in vivo* (109). Interestingly, a second MIF secretion peak was observed 8 h after the hypoxic trigger (59), indicating that within this intermediate time window MIF and CXCL12 may jointly act to drive EPC recruitment and neovascularisation (► Figure 1).

EPC recruitment and neovascularisation also represent important mechanisms driving the healing process of acute or chronic skin wounds. Accordingly, MIF was defined as a potential inducer of EPC mobilisation after flap operations. In flap patients, the number of circulating EPCs and serum levels of MIF but not CXCL12 serum levels was increased, especially in the patient group of free microvascular flaps. Also serum MIF and EPC levels correlated and MIF blockade, and to a lesser extent CXCL12 inhibition, partially reverted the chemotactic effect of patient serum on isolated human EPCs (110). The study also indicated that MIF-mediated EPC mobilisation is dependent on the degree of ischaemia (110).

As mentioned above, the neovascularisation potential of EPCs is in part due to the fact that these cells are carriers of numerous potent angiogenic/vasculogenic factors ('angiogenic cargo'). Prominent cargo factors are VEGF and thymosin- β 4, but interestingly also MIF and other chemokines (111, 112). The differential usage of these factors was recently further refined by *in vivo* implantation experiments, indicating that the MIF/chemokine receptor axis has an important role in differentiation towards an endothelial and SMC phenotype (113). As murine embryonic EPCs were previously found to induce blood vessel growth and cardioprotection under conditions of severe acute and chronic ischaemia in a mouse I/R model and a rat hind-limb ischaemia model, the above study confirms the therapeutic neovascularisation potential of EPCs in combination with selected sets of angiogenic chemokines/factors, including MIF (112, 113).

Acknowledgements

We thank Elisa Liehn, Heidi Noels, David Simons, Nancy Tuchscheerer and other doctoral researchers of the EuCAR program as well as numerous (inter)national collaborators and friends for helpful discussions over the past years. Our studies on the role of MIF and other chemokines in neovascularisation and atherosclerosis were supported by an IZKF Aachen grant of the Faculty of Medicine, RWTH Aachen University to J. B. and by Deutsche Forschungsgemeinschaft (DFG) grants BE1977/4-2/FOR 809 to J. B. and DFG-GRK1508/1 to J. B., Y. A., and M. S.

Conflicts of interest

None declared.

References

1. Charo IF, Ransohoff RM. The many roles of chemokines and chemokine receptors in inflammation. *N Engl J Med* 2006; 354: 610-621.
2. Laudanna C, Alon R. Right on the spot. Chemokine triggering of integrin-mediated arrest of rolling leukocytes. *Thromb Haemost* 2006; 95: 5-11.
3. Weber C, Schober A, Zerneck A. Chemokines: key regulators of mononuclear cell recruitment in atherosclerotic vascular disease. *Arterioscler Thromb Vasc Biol* 2004; 24: 1997-2008.
4. Murphy PM, Baggiolini M, Charo IF, et al. International union of pharmacology. XXII. Nomenclature for chemokine receptors. *Pharmacol Rev* 2000; 52: 145-176.
5. Koenen RR, Weber C. Chemokines: established and novel targets in atherosclerosis. *EMBO Mol Med* 2011; 3: 713-725.
6. Weber C. Chemokines take centre stage in vascular biology. *Thromb Haemost* 2007; 97: 685-687.
7. Humbert M, Morrell NW, Archer SL, et al. Cellular and molecular pathobiology of pulmonary arterial hypertension. *J Am Coll Cardiol* 2004; 43: 13S-24S.
8. Murdoch C, Finn A. Chemokine receptors and their role in vascular biology. *J Vasc Res* 2000; 37: 1-7.
9. Thelen M. Dancing to the tune of chemokines. *Nat Immunol* 2001; 2: 129-134.
10. von Hundelshausen P, Koenen RR, Sack M, et al. Heterophilic interactions of platelet factor 4 and RANTES promote monocyte arrest on endothelium. *Blood* 2005; 105: 924-930.
11. Koenen RR, von Hundelshausen P, Nesmelova IV, et al. Disrupting functional interactions between platelet chemokines inhibits atherosclerosis in hyperlipidemic mice. *Nat Med* 2009; 15: 97-103.
12. Moore BB, Arenberg DA, Addison CL, et al. CXC chemokines mechanism of action in regulating tumour angiogenesis. *Angiogenesis* 1998; 2: 123-134.

13. Strieter RM, Polverini PJ, Kunkel SL, et al. The functional role of the ELR motif in CXC chemokine-mediated angiogenesis. *J Biol Chem* 1995; 270: 27348-27357.
14. Belperio JA, Keane MP, Arenberg DA, et al. CXC chemokines in angiogenesis. *J Leukoc Biol* 2000; 68: 1-8.
15. Arenberg DA, Kunkel SL, Polverini PJ, et al. Inhibition of interleukin-8 reduces tumorigenesis of human non-small cell lung cancer in SCID mice. *J Clin Invest* 1996; 97: 2792-2802.
16. Takamori H, Oades ZG, Hoch OC, et al. Autocrine growth effect of IL-8 and GROalpha on a human pancreatic cancer cell line, Capan-1. *Pancreas* 2000; 21: 52-56.
17. Yoneda J, Kuniyasu H, Crispens MA, et al. Expression of angiogenesis-related genes and progression of human ovarian carcinomas in nude mice. *J Natl Cancer Inst* 1998; 90: 447-454.
18. Simonini A, Moscucci M, Muller DW, et al. IL-8 is an angiogenic factor in human coronary atherectomy tissue. *Circulation* 2000; 101: 1519-1526.
19. Peeters W, Hellings WE, de Kleijn DP, et al. Carotid atherosclerotic plaques stabilize after stroke: insights into the natural process of atherosclerotic plaque stabilisation. *Arterioscler Thromb Vasc Biol* 2009; 29: 128-133.
20. Gerszten RE, Garcia-Zepeda EA, Lim YC, et al. MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature* 1999; 398: 718-723.
21. Weber KS, von Hundelshausen P, Clark-Lewis I, et al. Differential immobilisation and hierarchical involvement of chemokines in monocyte arrest and transmigration on inflamed endothelium in shear flow. *Eur J Immunol* 1999; 29: 700-712.
22. Gu L, Okada Y, Clinton SK, et al. Absence of monocyte chemoattractant protein-1 reduces atherosclerosis in low density lipoprotein receptor-deficient mice. *Mol Cell* 1998; 2: 275-281.
23. Boring L, Gosling J, Cleary M, et al. Decreased lesion formation in CCR2^{-/-} mice reveals a role for chemokines in the initiation of atherosclerosis. *Nature* 1998; 394: 894-897.
24. Wakasugi K, Schimmel P. Two distinct cytokines released from a human aminoacyl-tRNA synthetase. *Science* 1999; 284: 147-151.
25. Wakasugi K, Slike BM, Hood J, et al. Induction of angiogenesis by a fragment of human tyrosyl-tRNA synthetase. *J Biol Chem* 2002; 277: 20124-20126.
26. Yang XL, Skene RJ, McRee DE, et al. Crystal structure of a human aminoacyl-tRNA synthetase cytokine. *Proc Natl Acad Sci U S A* 2002; 99: 15369-15374.
27. Howard OM, Dong HE, Yang D, et al. Histidyl-tRNA synthetase and asparaginyl-tRNA synthetase, autoantigens in myositis, activate chemokine receptors on T lymphocytes and immature dendritic cells. *J Exp Med* 2002; 196: 781-791.
28. Yang D, Chertov O, Bykovskaia SN, et al. Beta-defensins: linking innate and adaptive immunity through dendritic and T cell CCR6. *Science* 1999; 286: 525-528.
29. Alcamí A. Viral mimicry of cytokines, chemokines and their receptors. *Nat Rev Immunol* 2003; 3: 36-50.
30. Noels H, Bernhagen J, Weber C. Macrophage migration inhibitory factor: a noncanonical chemokine important in atherosclerosis. *Trends Cardiovasc Med* 2009; 19: 76-86.
31. Murphy PM. Viral exploitation and subversion of the immune system through chemokine mimicry. *Nat Immunol* 2001; 2: 116-122.
32. Andersson U, Tracey KJ. HMGB1 is a therapeutic target for sterile inflammation and infection. *Annu Rev Immunol* 2011; 29: 139-162.
33. Lotze MT, Tracey KJ. High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nat Rev Immunol* 2005; 5: 331-342.
34. Schiraldi M, Raucci A, Munoz LM, et al. HMGB1 promotes recruitment of inflammatory cells to damaged tissues by forming a complex with CXCL12 and signalling via CXCR4. *J Exp Med* 2012; 209: 551-563.
35. David JR. Delayed hypersensitivity in vitro: its mediation by cell-free substances formed by lymphoid cell-antigen interaction. *Proc Natl Acad Sci USA* 1966; 56: 72-77.
36. Degryse B, de Virgilio M. The nuclear protein HMGB1, a new kind of chemokine? *FEBS Lett* 2003; 553: 11-17.
37. Calandra T, Roger T. Macrophage migration inhibitory factor: a regulator of innate immunity. *Nat Rev Immunol* 2003; 3: 791-800.
38. Morand EF, Leech M, Bernhagen J. MIF: a new cytokine link between rheumatoid arthritis and atherosclerosis. *Nat Rev Drug Discov* 2006; 5: 399-410.
39. Noels H, Bernhagen J, Weber C. MIF in atherosclerosis. In: Bucala R, editor. *The MIF Handbook*. World Scientific Publishing, Hongkong; 2012.
40. Leng L, Metz CN, Fang Y, et al. MIF signal transduction initiated by binding to CD74. *J Exp Med* 2003; 197: 1467-1476.
41. Shi X, Leng L, Wang T, et al. CD44 is the signalling component of the macrophage migration inhibitory factor-CD74 receptor complex. *Immunity* 2006; 25: 595-606.
42. Bernhagen J, Krohn R, Lue H, et al. MIF is a noncognate ligand of CXC chemokine receptors in inflammatory and atherogenic cell recruitment. *Nat Med* 2007; 13: 587-596.
43. Schwartz V, Lue H, Kraemer S, et al. A functional heteromeric MIF receptor formed by CD74 and CXCR4. *FEBS Lett* 2009; 583: 2749-2757.
44. Liu Y, Shaw SK, Ma S, et al. Regulation of leukocyte transmigration: cell surface interactions and signalling events. *J Immunol* 2004; 172: 7-13.
45. Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. *Nat Rev Immunol* 2007; 7: 803-815.
46. Danese S, Dejana E, Fiocchi C. Immune regulation by microvascular endothelial cells: directing innate and adaptive immunity, coagulation, and inflammation. *J Immunol* 2007; 178: 6017-6022.
47. Donnelly SC, Bucala R. Macrophage migration inhibitory factor: a regulator of glucocorticoid activity with a critical role in inflammatory disease. *Mol Med Today* 1997; 3: 502-507.
48. Lolis E, Bucala R. Therapeutic approaches to innate immunity: severe sepsis and septic shock. *Nat Rev Drug Discov* 2003; 2: 635-645.
49. Mitchell RA, Bucala R. Tumour growth-promoting properties of macrophage migration inhibitory factor. *Semin Cancer Biol* 2000; 10: 359-366.
50. Bernhagen J, Calandra T, Mitchell RA, et al. MIF is a pituitary-derived cytokine that potentiates lethal endotoxaemia. *Nature* 1993; 365: 756-759.
51. Calandra T, Spiegel LA, Metz CN, et al. Macrophage migration inhibitory factor is a critical mediator of the activation of immune cells by exotoxins of Gram-positive bacteria. *Proc Natl Acad Sci USA* 1998; 95: 11383-11388.
52. Bozza M, Satskar AR, Lin G, et al. Targeted disruption of migration inhibitory factor gene reveals its critical role in sepsis. *J Exp Med* 1999; 189: 341-346.
53. Calandra T, Echtenacher B, Roy DL, et al. Protection from septic shock by neutralisation of macrophage migration inhibitory factor. *Nat Med* 2000; 6: 164-170.
54. Veillat V, Carli C, Metz CN, et al. Macrophage migration inhibitory factor elicits an angiogenic phenotype in human ectopic endometrial cells and triggers the production of major angiogenic factors via CD44, CD74, and MAPK signalling pathways. *J Clin Endocrinol Metab* 2010; 95: E403-412.
55. Baugh JA, Chitnis S, Donnelly SC, et al. A functional promoter polymorphism in the macrophage migration inhibitory factor (MIF) gene associated with disease severity in rheumatoid arthritis. *Genes Immun* 2002; 3: 170-176.
56. Burger-Kentischer A, Goebel H, Seiler R, et al. Expression of macrophage migration inhibitory factor in different stages of human atherosclerosis. *Circulation* 2002; 105: 1561-1566.
57. Shimizu T, Nishihira J, Watanabe H, et al. Macrophage migration inhibitory factor is induced by thrombin and factor Xa in endothelial cells. *J Biol Chem* 2004; 279: 13729-13737.
58. Schober A, Bernhagen J, Thiele M, et al. Stabilisation of atherosclerotic plaques by blockade of macrophage migration inhibitory factor after vascular injury in apolipoprotein E-deficient mice. *Circulation* 2004; 109: 380-385.
59. Simons D, Grieb G, Hristov M, et al. Hypoxia-induced endothelial secretion of macrophage migration inhibitory factor and role in endothelial progenitor cell recruitment. *J Cell Mol Med* 2011; 15: 668-678.
60. Chen L, Yang G, Zhang X, et al. Induction of MIF expression by oxidized LDL via activation of NF- κ B in vascular smooth muscle cells. *Atherosclerosis* 2009; 207: 428-433.
61. Schrans-Stassen BH, Lue H, Sonnemans DG, et al. Stimulation of vascular smooth muscle cell migration by macrophage migration inhibitory factor. *Antioxid Redox Signal* 2005; 7: 1211-1216.
62. Lin SG, Yu XY, Chen YX, et al. De novo expression of macrophage migration inhibitory factor in atherogenesis in rabbits. *Circ Res* 2000; 87: 1202-1208.
63. Cheng Q, McKeown SJ, Santos L, et al. Macrophage migration inhibitory factor increases leukocyte-endothelial interactions in human endothelial cells via promotion of expression of adhesion molecules. *J Immunol* 2010; 185: 1238-1247.
64. Amin MA, Haas CS, Zhu K, et al. Migration inhibitory factor up-regulates vascular cell adhesion molecule-1 and intercellular adhesion molecule-1 via Src, PI3 kinase, and NF κ B. *Blood* 2006; 107: 2252-2261.

65. Chen Z, Sakuma M, Zago AC, et al. Evidence for a role of macrophage migration inhibitory factor in vascular disease. *Arterioscler Thromb Vasc Biol* 2004; 24: 709-714.
66. Libby P. Inflammation in atherosclerosis. *Nature* 2002; 420: 868-874.
67. Schmeisser A, Marquetant R, Illmer T, et al. The expression of macrophage migration inhibitory factor 1alpha (MIF 1alpha) in human atherosclerotic plaques is induced by different proatherogenic stimuli and associated with plaque instability. *Atherosclerosis* 2005; 178: 83-94.
68. de Winther MP, Kanters E, Kraal G, et al. Nuclear factor kappaB signalling in atherogenesis. *Arterioscler Thromb Vasc Biol* 2005; 25: 904-914.
69. Schweitzer K, Bozko PM, Dubiel W, et al. CSN controls NF-kappaB by deubiquitinylation of IkappaBalpha. *Embo J* 2007; 26: 1532-1541.
70. Kleemann R, Hausser A, Geiger G, et al. Intracellular action of the cytokine MIF to modulate AP-1 activity and the cell cycle through Jab1. *Nature* 2000; 408: 211-216.
71. Pan JH, Sukhova GK, Yang JT, et al. Macrophage migration inhibitory factor deficiency impairs atherosclerosis in low-density lipoprotein receptor-deficient mice. *Circulation* 2004; 109: 3149-3153.
72. Burger-Kentischer A, Gobel H, Kleemann R, et al. Reduction of the aortic inflammatory response in spontaneous atherosclerosis by blockade of macrophage migration inhibitory factor (MIF). *Atherosclerosis* 2006; 184: 28-38.
73. Kraemer S, Lue H, Zerneck A, et al. MIF-chemokine receptor interactions in atherogenesis are dependent on an N-loop-based 2-site binding mechanism. *FASEB J* 2011; 25: 894-906.
74. Herder C, Illig T, Baumert J, et al. Macrophage migration inhibitory factor (MIF) and risk for coronary heart disease: results from the MONICA/KORA Augsburg case-cohort study, 1984-2002. *Atherosclerosis* 2008; 200: 380-388.
75. Tereshchenko IP, Petrakova J, Mrazek F, et al. The macrophage migration inhibitory factor (MIF) gene polymorphism in Czech and Russian patients with myocardial infarction. *Clin Chim Acta* 2009; 402: 199-202.
76. Makino A, Nakamura T, Hirano M, et al. High plasma levels of macrophage migration inhibitory factor are associated with adverse long-term outcome in patients with stable coronary artery disease and impaired glucose tolerance or type 2 diabetes mellitus. *Atherosclerosis* 2010; 213: 573-578.
77. Muller II, Muller KA, Schonleber H, et al. Macrophage migration inhibitory factor is enhanced in acute coronary syndromes and is associated with the inflammatory response. *PLoS One* 2012; 7: e38376.
78. Conway EM, Collen D, Carmeliet P. Molecular mechanisms of blood vessel growth. *Cardiovasc Res* 2001; 49: 507-521.
79. Buschmann I, Schaper W. Arteriogenesis versus angiogenesis: two mechanisms of vessel growth. *News Physiol Sci* 1999; 14: 121-125.
80. Risau W, Flamme I. Vasculogenesis. *Annu Rev Cell Dev Biol* 1995; 11: 73-91.
81. Asahara T, Murohara T, Sullivan A, et al. Isolation of putative progenitor endothelial cells for angiogenesis. *Science* 1997; 275: 964-967.
82. Ingram DA, Caplice NM, Yoder MC. Unresolved questions, changing definitions, and novel paradigms for defining endothelial progenitor cells. *Blood* 2005; 106: 1525-1531.
83. Ingram DA, Mead LE, Tanaka H, et al. Identification of a novel hierarchy of endothelial progenitor cells using human peripheral and umbilical cord blood. *Blood* 2004; 104: 2752-2760.
84. Rehman J, Li J, Orschell CM, et al. Peripheral blood "endothelial progenitor cells" are derived from monocyte/macrophages and secrete angiogenic growth factors. *Circulation* 2003; 107: 1164-1169.
85. Simons M, Ware JA. Therapeutic angiogenesis in cardiovascular disease. *Nat Rev Drug Discov* 2003; 2: 863-871.
86. Tuchscheerer N. The ligands of CXCR4 in vascularisation [Dissertation thesis]. Aachen: RWTH Aachen University; 2012.
87. Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. *Nature* 2011; 473: 298-307.
88. Yancopoulos GD, Davis S, Gale NW, et al. Vascular-specific growth factors and blood vessel formation. *Nature* 2000; 407: 242-248.
89. Chesney J, Metz C, Bacher M, et al. An essential role for macrophage migration inhibitory factor (MIF) in angiogenesis and the growth of a murine lymphoma. *Mol Med* 1999; 5: 181-191.
90. Wilson JM, Coletta PL, Cuthbert RJ, et al. Macrophage migration inhibitory factor promotes intestinal tumorigenesis. *Gastroenterology* 2005; 129: 1485-1503.
91. Winner M, Meier J, Zierow S, et al. A novel, macrophage migration inhibitory factor suicide substrate inhibits motility and growth of lung cancer cells. *Cancer Res* 2008; 68: 7253-7257.
92. Rendon BE, Roger T, Teneng I, et al. Regulation of human lung adenocarcinoma cell migration and invasion by macrophage migration inhibitory factor. *J Biol Chem* 2007; 282: 29910-29918.
93. Brock SE, Rendon BE, Yaddanapudi K, et al. Negative Regulation of AMP-activated Protein Kinase (AMPK) Activity by Macrophage Migration Inhibitory Factor (MIF) Family Members in Non-small Cell Lung Carcinomas. *J Biol Chem* 2012; 287: 37917-37925.
94. Rendon BE, Willer SS, Zundel W, et al. Mechanisms of macrophage migration inhibitory factor (MIF)-dependent tumour microenvironmental adaptation. *Exp Mol Pathol* 2009; 86: 180-185.
95. Coleman AM, Rendon BE, Zhao M, et al. Cooperative regulation of non-small cell lung carcinoma angiogenic potential by macrophage migration inhibitory factor and its homolog, D-dopachrome tautomerase. *J Immunol* 2008; 181: 2330-2337.
96. Baugh JA, Gantier M, Li L, et al. Dual regulation of macrophage migration inhibitory factor (MIF) expression in hypoxia by CREB and HIF-1. *Biochem Biophys Res Commun* 2006; 347: 895-903.
97. Copple BL, Bai S, Burgoon LD, et al. Hypoxia-inducible factor-1alpha regulates the expression of genes in hypoxic hepatic stellate cells important for collagen deposition and angiogenesis. *Liver Int* 2011; 31: 230-244.
98. Fu H, Luo F, Yang L, et al. Hypoxia stimulates the expression of macrophage migration inhibitory factor in human vascular smooth muscle cells via HIF-1alpha dependent pathway. *BMC Cell Biol* 2010; 11: 66.
99. Amin MA, Volpert OV, Woods JM, et al. Migration inhibitory factor mediates angiogenesis via mitogen-activated protein kinase and phosphatidylinositol kinase. *Circ Res* 2003; 93: 321-329.
100. Lue H, Kapurniotu A, Fingerle-Rowson G, et al. Rapid and transient activation of the ERK MAPK signalling pathway by macrophage migration inhibitory factor (MIF) and dependence on JAB1/CSN5 and Src kinase activity. *Cell Signal* 2006; 18: 688-703.
101. Mitchell RA, Metz CN, Peng T, et al. Sustained mitogen-activated protein kinase (MAPK) and cytoplasmic phospholipase A2 activation by macrophage migration inhibitory factor (MIF). Regulatory role in cell proliferation and glucocorticoid action. *J Biol Chem* 1999; 274: 18100-18106.
102. Liehn EA, Merx MW, Postea O, et al. Ccr1 deficiency reduces inflammatory remodelling and preserves left ventricular function after myocardial infarction. *J Cell Mol Med* 2008; 12: 496-506.
103. Liehn EA, Tuchscheerer N, Kanzler I, et al. Double-edged role of the CXCL12/CXCR4 axis in experimental myocardial infarction. *J Am Coll Cardiol* 2011; 58: 2415-2423.
104. Koga K, Kenessey A, Powell S, et al. Macrophage migration inhibitory factor provides cardioprotection during ischaemia/reperfusion by reducing oxidative stress. *Antioxid Redox Signal* 2010; 14: 1191-1202.
105. Miller EJ, Li J, Leng L, et al. Macrophage migration inhibitory factor stimulates AMP-activated protein kinase in the ischaemic heart. *Nature* 2008; 451: 578-582.
106. Qi D, Hu X, Wu X, et al. Cardiac macrophage migration inhibitory factor inhibits JNK pathway activation and injury during ischaemia/reperfusion. *J Clin Invest* 2009; 119: 3807-3816.
107. Luedike P, Hendgen-Cotta UB, Sobierajski J, et al. Cardioprotection through S-nitrosylation of macrophage migration inhibitory factor. *Circulation* 2012; 125: 1880-1889.
108. Ceradini DJ, Gurtner GC. Homing to hypoxia: HIF-1 as a mediator of progenitor cell recruitment to injured tissue. *Trends Cardiovasc Med* 2005; 15: 57-63.
109. Ceradini DJ, Kulkarni AR, Callaghan MJ, et al. Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. *Nat Med* 2004; 10: 858-864.
110. Grieb G, Piatkowski A, Simons D, et al. Macrophage migration inhibitory factor is a potential inducer of endothelial progenitor cell mobilisation after flap operation. *Surgery* 2012; 151: 268-277.
111. Kupatt C, Bock-Marquette I, Boekstegers P. Embryonic endothelial progenitor cell-mediated cardioprotection requires Thymosin beta4. *Trends Cardiovasc Med* 2008; 18: 205-210.
112. Kupatt C, Horstkotte J, Vlastos GA, et al. Embryonic endothelial progenitor cells expressing a broad range of proangiogenic and remodelling factors enhance vascularisation and tissue recovery in acute and chronic ischaemia. *FASEB J* 2005; 19: 1576-1578.
113. Kanzler I, Tuchscheerer N, Steffens G, et al. Differential roles of angiogenic chemokines in endothelial progenitor cell-induced angiogenesis. *Basic Res Cardiol*